

BREW PLATE



GOLD KIT INSTRUCTIONS



PRECAUTIONS

1. **Storage of Brew Plates:** Always store or keep Brew Plates lid down (upside down, similar to how they are stored in the kit) to prevent water condensation from collecting on the surface of the media.
2. **Do not place any material provided in the kit directly into brew.**

PRE-SAMPLING CLEAN UP

1. **Wash hands thoroughly.**
2. **Work area:** Locate table top or any flat surface on which to work. Immediately before working with your Brew Plates clean this area with your cleaning agent of choice (e.g. 10% Bleach, Lysol spray or wipes, Alcohol wipes) and allow area to dry.
3. **Sampling site:** Wipe down with a cleaning agent or flame any valve or equipment that the liquid sample may come into contact with during collection. If the equipment or a site will be tested for contamination, do not pre-clean before taking a sample.

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INCUBATION OF PLATES AND TUBES:

1. When all the Brew Plate agar zones are inoculated with your liquid, place the plates face down into a Ziploc bag provided, and without sealing the bag, place it into the Brew Plate Incubator set at 30 degrees Celsius. This bag will prevent the plates from drying from the ventilation from the fan inside the incubator, and the bacteria and/or yeast will still have access to oxygen to grow.
2. Place the inoculated HLP tubes into the Brew Plate Incubator set at 30 degrees Celsius.
3. Allow for up to 5 days to incubate.

[Note: Incubation time may vary depending on the type of microbial contaminant: aerobic bacteria (1 to 2 days), yeast (1 to 2 days), and anaerobic bacteria (2 to 5 days in HLP tubes).]

4. After the recommended incubation time has passed, remove the Brew Plates from the incubator and examine for growth on the surface of the agar. Remove the HLP tubes from the incubator and examine for growth in the semi-solid medium looking for droplet shaped bacterial growth.
5. If wild yeast or bacteria grow on the media or in the HLP tube and you choose to have the microbe identified, go to www.BrewPlate.com to place an order.

[Note: Brew Plate offers one (1) free microbe identification per purchased kit or plates.]

6. Before you place an order, you need to create a user account. To place an order, use your login, password, and Brew Plate kit number found on the box to establish the on-line order. Next, indicate the number of microbes you wish identified. Please provide the Media Code (see list below) that corresponds to the location of the microbe(s) you want identified and provide a brief description of the microbe for further clarity if more than one microbe is detected. Payments are processed using most major credit cards. The provided 'payment confirmation code' will be needed for 'mailing the brew plate.'
7. Send the plates and/or HLP tubes to Brew Plate Laboratories for identification.

Mailing the Brew Plate:

- a. Label the bottom of the Brew Plate and/or the HLP tube with both the
 - i. Brew Plate Kit number provided on the box and
 - ii. Payment confirmation code.
- b. If you wish to identify the microbe you want identified, line up the line on the plate lid to the line on the plate bottom and with a 'Sharpie' marker, circle the colony or colonies on the top of the plate lid making sure it aligns with the colony when looking through the lid to the agar.
- c. Wrap the plate(s) in clean plastic wrap, then place them into the provided return envelope.
- d. Place the plate into the provided mailing envelope with the Brew Plate return address and mail the envelope to Brew Plate Laboratories.
- e. To see your test results, log back into your account at www.BrewPlate.com.

Mailing Address:

Brew Plate Laboratories
2439 Kuser Road
Hamilton, NJ 08690

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TESTING LIQUID SAMPLES:

Collection of Liquid sample

1. Pour or collect at least 2 ml of liquid to be tested in the sterile tubes provided.
2. Keep sample tubes capped until you are ready to apply the sample to the plates.

Applying sample to the Brew Plates

1. Remove the number of Brew Plates from the sterile packaging that you intend to use. Reseal the plastic packaging containing the unused plates with a clean clip (e.g. paper clip wiped with cleaning agent). Resealing will keep the unused plates sterile and prevent dehydration.
2. Place the collection tube containing the liquid to be tested in the stand provided.
3. Remove the cap from the collection tube and place on the clean surface bottom up.
4. Remove a sterile swab from the kit. Hold the swab by the handle being careful not to touch the tip or place it on unclean surfaces.
5. Dip the swab into the liquid inside collection tube. Gently place the inoculated swab on top of a Brew Plate agar in the center of the zone. Move the swab across the surface of the agar in a zig-zag manner. Repeat until the liquid has been absorbed into the agar. Be sure to inoculate as much of the agar surface as possible and be careful not to turn the swab while spreading. Turn the swab 90 degrees (for quad plates) or 180 degrees (for bi-plates) and spread onto the next quadrant surface until all quadrants are covered. Repeat step 7 to apply the sample to the other Brew Plates.

Applying sample into the HLP Tubes

1. Carefully and cleanly remove the cap of the provided HLP semi-solid media culture tube.
2. Remove a sterile loop from the kit. Hold the loops by the handle being careful not to touch the circular opening or place it on unclean surfaces.
3. Dip the loop into the liquid inside collection tube. Make sure the loop has the full amount of liquid.
4. Place the inoculated loop $\frac{3}{4}$ of the way down into the media.
5. Remove the inoculating loop and recap the HLP culture tube.

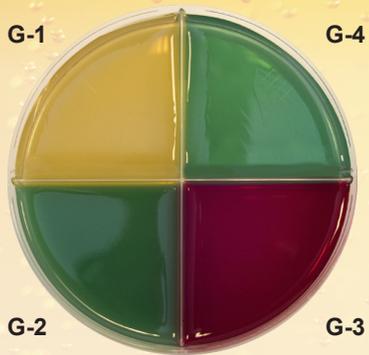
TESTING SURFACES AND EQUIPMENT:

Collection of sample:

1. Use a sterile swab to sample the surfaces of tanks or other equipment for contamination. If the surface is dry, lightly moisten the swab with the sterile water provided in the kit before taking the sample.
2. Swipe and roll the swab over the area of interest.

Applying sample to the Brew Plates

1. Remove the number of Brew Plates from the sterile packaging that you intend to use. Reseal the plastic packaging containing the unused plates with a clean clip (e.g. paper clip wiped with cleaning agent). Resealing will keep the unused plates sterile and prevent dehydration.
2. Gently swipe and roll the swab over the surface of the Brew Plate zone starting at the center of the zone. Be sure to inoculate as much of the zone surface as possible.
3. Repeat steps 1-3 to apply the sample to the other Brew Plate zone.



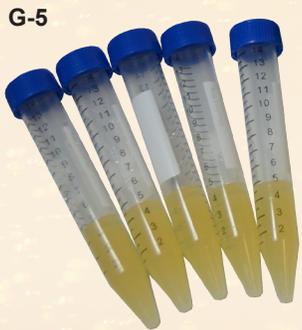
BREW PLATE – GOLD KIT – Quad-Plate

Quadrant I: Lin's Cupric Sulfate Media (LCSM)(Clear color)
 Microbes Isolated: Non-Saccharomyces wild yeast
 Sequence ID Media Code: **G-1**

Quadrant II: Lee's Multi-Differential Agar (LMDA)(Green color) with Cycloheximide
 Microbes Isolated: Bacteria with color and morphology differential
 Sequence ID Media Code: **G-2**

Quadrant III: Lin's Wild Yeast Media (LWYM)(Purple/Dark Red color)
 Microbes Isolated: Saccharomyces wild yeast
 Sequence ID Media Code: **G-3**

Quadrant IV: Wallerstein Nutrient Media (WLN)(Blue-Green color)
 Microbes Isolated: Yeast and Bacteria with color and morphology differential
 Sequence ID Media Code: **G-4**



BREW PLATE – GOLD KIT – HLP Tube

Tube: Hsu's Lactobacillus/Pediococcus semi-solid media (HLP)
 Microbes Isolated: Lactobacillus and Pediococcus bacteria
 Sequence ID Media Code: **G-5**

Disposal of Kit Contents

All non-contaminated plastic and cardboard components may be recycled. Used plates that are not sent to Brew Plate Laboratories can be discarded with your pitching yeast disposable materials. It is best to wrap the plates with plastic wrap before discarding to prevent yeast or bacterial growth on the plate from being a potential source of further contamination.